REVISED STRUCTURE OF CERIFERIC ACID

Yoko NAYA*, Fumiko MIYAMOTO, Keiko KISHIDA, Takenori KUSUMI[†], Hiroshi KAKISAWA[†], and Koji NAKANISHI Suntory Institute for Bioorganic Research**, Shimamoto-cho, Mishima-gun, Osaka 618 Department of Chemistry, The University of Tsukuba, Niihari-gun, Sakura-mura, Ibaraki 300-31

The structure of ceriferic acid, a scale insect sesterterpene secretion, has been revised from 10 to 5. Naturally occurring ceriferol has been correlated with ceriferic acid.

We recently reported the structures of a series of 14-membered monocyclic sesterterpenoids isolated from the secretion of a Japanese scale insect Ceroplastes ceriferus Anderson (Coccidae). 1,2 Eight of these possess the 2-t/6c/10-t skeletal framework as exemplified by cericerol-I ($\underline{1}$),[α] $_{D}^{27}$ -84.1°, for which the absolute configuration at C-14 was established by chemical correlations; two minor congenors possibly have 2-t/6-c/10-c and 2-t/6-t/10-t skeletons.

The arrangement of annular double bonds in these compounds is in contrast to that of albocerol (2) which was isolated from the Mexican species C. albolineatus by Veloz et al. 3 It should be noted that the skeletons of $\frac{1}{2}$ and $\frac{2}{2}$ are derived, respectively, from two opposing cyclization modes of the biogenetic precursor geranylfarnesyl pyrophosphate (3) and (4). It was thus quite remarkable that spectral data of ceriferic acid isolated from the Japanese species C. ceriferus (collected in Tokyo area) led to the albocerol- or Mexican-type structure (10); 4 it even implied necessity for a taxonomic reinvestigation. In the following, however, we show that ceriferic acid should after all be represented by the cericerol-I type structure (5).

The evidence leading to the revised structure is as follows: 1) Ceriferic acid showed no CD Cotton effect. Therefore, the α,β -unsaturated carboxylic acid group, which from $^{1}\text{H-}$ and $^{13}\text{C-NMR}$ data is clearly part of a cis

 $(\underline{1}): R=CH_2OH$ (cericerol-I)

(2) (albocerol)

 $(11): R=CH_3$

(3)

R

10

(5): $R=C0_2H$ (ceriferic acid)

(6): $R=C0_2CH_3$

 $\begin{bmatrix} & & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & \\ & & & \\ & \\ & & \\ & &$

 $(\underline{4})$

 $(\underline{7}): R=CH_2OH$ (ceriferol)

 $(8): R=CH_2OTs$

 $(\underline{9}): R=CH_3$

Table 1. $^{13}\text{C-NMR}$ chemical shifts of methyl ceriferate (6), ceriferol (7), " α -cericerene" (9), and cericerene (11) in CDCl $_3$. Peak assignments are based on measurements of T_1 and $^{13}\text{C-NOE}^1$ in addition to the more conventional techniques.

C- <u>a</u> tom	<u>6</u>	7	<u>9</u>	11
C-1	29.8 ^a	29.6 ^a	29.4 ^a	30.5 ^a
C-2	125.3 ^b	125.0 ^b	125.2 ^b	125.1 ^b
C-3	133.7 ^C	133.2 ^c	133.8 ^C	134.1 ^C
C-4	31.2 ^a	29.9 ^a	30.4 ^a	31.1 ^a
C-5	26.9	27.2	31.0 ^a	31.4 ^a
C-6	142.4	127.6	124.9 ^b	125.1 ^b
C-7	131.3 ^d	137.6	133.1 ^c	132.9 ^c
C-8	36.0	35.8	35.8	36.2
C-9	26.0	24.6 ^d	24.5	24.6
C-10	125.4 ^b	125.0 ^b	124.9 ^b	125.0 ^b
C-11	132.9 ^C	133.1 ^C	132.9 ^C	133.0 ^C
C-12	40.3	30.2	40.2	40.3
C-13	24.5	24.5 ^d	24.5	24.6
C-14	49.9	46.4	46.6	44.6
C-15	136.4	136.8	137.1	153.0
C-16	123.4	123.5	123.7	33.7
C-17	26.9	26.9	26.8	26.6
C-18	124.6	124.8 ^b	124.5	124.6 ^b
C-19	131.1 ^d	131.2	131.0	131.3
C-20	25.7	25.6	25.6	25.7
C-21	17.7	17.7	17.7	17.8
C-22	12.3	12.0	12.0	108.9
C-23	15.6 ^e	15.6 ^e	15.6 ^d	15.6 ^d
C-24	168.4	66.6	22.4	22.5
C-25	15.3 ^e	15.4 ^e	15.4 ^d	15.5 ^d
-соо <u>с</u> н ₃	51.3			

a-e) Assignments denoted by same alphabet are interchangeable.

- skeleton framework, $\frac{4}{}$ must be remote from the sole chiral center at C-14; if it were as in $\frac{10}{}$, the CD should have shown a Cotton effect at the λ_{\max} of ceriferic acid. Of the two 14-membered skeletal possibilities, $\frac{5}{}$ (or $\frac{1}{}$) and $\frac{10}{}$, it is only structure 5 which satisfies the two criteria underscored above.
- 2) Reduction of methyl ceriferate ($\underline{6}$) with LiAlH $_4$ gave "ceriferol" ($\underline{7}$), which was further converted into the tosylate ($\underline{8}$) by careful treatment with TsCl/py at 0 °C. Tosylate $\underline{8}$ with its strongly absorbing chromophore also lacked a CD Cotton effect, thus confirming the deductions described above.
- 3) Acid treatment of tosylate $\underline{8}$ simply resulted in hydrolysis to alcohol $\underline{7}$. In contrast, acid treatment of the tosylate derived from cericerol-I $\underline{1}$ underwent transannular cyclization to a bicyclocericerene.
- 4) The neutral fraction of *C. ceriferus* collected around Kyoto in 1979 has given, in addition to cericerol-I $\frac{1}{2}$, an alcohol, $\left[\alpha\right]_{D}^{24}$ -83.5° (c, 1.33), the physical constants (MS, NMR, IR, and rotation) of which were identical with those of allyl alcohol $\frac{7}{2}$ (ceriferol). Furthermore, the acidic fraction gave ceriferic acid, $\left[\alpha\right]_{D}^{26}$ -97.6° (c, 1.02); these findings are in line with the taxonomical identify of the Tokyo and Kyoto species.
- 5) The 13 C-NMR data of ceriferic acid $\underline{5}$ derivatives are in good agreement with cericerol-I $\underline{1}$ derivatives, a further support for the skeletal identity. This is exemplified by a comparison of the data for hydrocarbons $(\underline{9})$, $[\alpha]_D^{19}$ -28.8° (c, 0.24) (derived from tosylate $\underline{8}$ by LiAlH₄ treatment) and $(\underline{11})_D^1$ $[\alpha]_D^{27}$ -48.3° (c, 0.48) (derived from cericerol-I) (Table 1). The data for methyl ceriferate $\underline{6}$ and ceriferol 7 given in Table 1 are in accord with the structural variations.

Finally, in analogy with other cericerol-I $\underline{1}$ derivatives, we assign an R-configuration (or " β " as depicted) to the C-14 of ceriferic acid 5.

REFERENCES

- 1) F. Miyamoto, H. Naoki, T. Takemoto, and Y. Naya, Tetrahedron, 35, 1913 (1979).
- 2) F. Miyamoto, H. Naoki, Y. Naya, and K. Nakanishi, Tetrahedron in press.
- 3) R. Veloz, L. Quijano, J.S. Calderon, and T. Rios, J.C.S. Chem. Commun., 1975, 191.
- 4) T. Kusumi, T. Kinoshita, K. Fujita, and H. Kakisawa, Chem. Lett., 1979, 1129.
- 5) To be published elsewhere.
- ** Previous name: The Institute of Food Chemistry